

Paediatric Cardiomyopathies EMCHC Investigation Guideline

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| Staff relevant to: | Medical staff within EMCHC, Cardiac PICU services and wider EMCH Network |
| Approval date: | August 2023 |
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| Written by: | Prof F Bu'Lock, Dr LJE Maddocks (Chemical Pathology), Dr S Shebani |
| Trust Ref: | C195/2016 |

Related Guidelines and Policies:

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| C39/2016 | Muscle and Skin Biopsy UHL Childrens Medical Guideline |
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1. Introduction

Cardiomyopathies can be:

- Dilated
- Hypertrophic
- Restrictive
- Left ventricular non-compaction
- Arrhythmogenic Right Ventricle

By far the commonest to present to paediatric cardiology are dilated and hypertrophic cardiomyopathies. Presentations are variable, causes are numerous and, in many cases, rare. The diagnosis is made and confirmed on ECG and echocardiogram. The rationale for identifying the causes of cardiomyopathies is to try to:

- Identify a possible condition
- Confirm the cause.
- Provide any condition specific treatments / prognosis / advice as appropriate

This guideline is for the use of medical staff to summarise the basic investigations required to identify a possible cause of cardiomyopathy and provide direction towards confirmatory tests.

2. Screening Tests: Please note several tests require special measures - read through the list and make sure you have everything necessary before bleeding the patient.

Table 1 Screening Tests

| 1st line screening tests | Sample Requirements | Date Sent | Opted out / why | Test Result & date |
|--|--|-----------|-----------------|--------------------|
| | (Bottles, special requirements, etc) | | | |
| Blood | | | | |
| Full blood count | 0.5ml EDTA | | | |
| Urea and electrolytes Liver function tests Bone profile Magnesium CK Uric Acid Cholesterol, triglyceride Creatinine Kinase | 1.2ml LiHep | | | |
| Coagulation screen | Filled Citrate | | | |
| Blood gas Blood glucose Bicarbonate Chloride Anion gap Lactate | | | | |
| Ketones | Ketostix | | | |
| Ammonia | 1ml LiHep, on ice, to lab within 30 minutes | | | |
| Lactate (if no facilities for a blood gas) | 1ml Fluoride, on ice, to lab within 30 minutes | | | |
| Pyruvate: please check comments from Biochemistry below | Special bottle, call lab on 16559 | | | |
| Free fatty acids | Fluoride, ideally hypoglycaemic sample | | | |
| Amino Acids | 0.5ml LiHep | | | |
| Acylcarnitine profile | 0.5ml LiHep | | | |
| Thyroid function | 1.2 ml LiHep | | | |
| Blood cultures | | | | |
| Viral PCR: <i>Enterovirus, echovirus, coxsackie virus, Parvovirus B19, consider HIV</i> | 1.2ml EDTA | | | |
| ASOT Rheumatic fever | 0.5ml serum | | | |
| Vitamin D | 0.5ml LiHep | | | |
| Selenium | 0.5ml serum | | | |
| Thiamine (Vit B1) | 1ml EDTA, protect from light, to lab within 30 minutes | | | |
| Urine | | | | |
| Amino acids | 1ml plain bottle | | | |
| Organic acids | 2ml plain bottle | | | |
| Urine Mucopolysaccharide Screen (MPS I, II, VI) Oligosaccharides Sialic Acid Glycosaminoglycans GAG electrophoresis | 5ml plain bottle, request MPS screen | | | |
| Ketones | Dipstick | | | |
| Glucose | Dipstick | | | |
| Reducing substances | 5ml plain bottle | | | |
| NB: A full set of 1st line blood investigations requires: 3 EDTA, 4 Lithium heparin, 1 citrate, 1 fluoride and 2 serum bottles and One blood gas | | | | |

| 2 nd line testing | Sample requirements | Date sent | Opted out / why | Result & date |
|--|---|-----------|-----------------|---|
| Muscle and Skin Biopsy for mitochondrial disorders | | | | |
| Histology | See muscle biopsy instructions | | | |
| Electron microscopy | | | | |
| Enzyme assays | | | | |
| Mitochondrial DNA analysis | | | | |
| Blood | | | | |
| Cardiolipin Levels (Barth Syndrome) | See proforma and letter below. EDTA 1-3ml | | | Monolysocardiolipin /cardiolipin phospholipid ratio |
| White cell enzymes <i>For lysosomal storage disorders (MPS II & III), if abnormal urinary oligosaccharides, GAG's, elevated CK or ALT/AST</i> | EDTA 5ml, to lab before 1pm, Mon-Thurs | | | |
| Transferrin and Apolipoprotein isoforms <i>if Congenital Disorder of Glycosylation suspected</i> | 1ml Clotted | | | |
| Phytanic Acid <i>if Refsum Disease suspected</i> | Call lab on 16559 | | | |
| Fabry Disease <i>Alpha glucosidase</i> | 1ml EDTA blood, request Fabry Screen | | | |
| Pompe's Screen <i>Acid-alpha glucosidase</i> | 1ml EDTA blood, request Pompe Screen | | | |
| Brain MRI | | | | |
| Suspected mitochondrial disorders | | | | |

NB: Testing for specific genetic conditions should be on the basis of the eligibility criteria according to the national genomic test directory as advised by Professor Pradeep Vasudevan Consultant Clinical Geneticist. Please see the link below.

<https://www.england.nhs.uk/wp-content/uploads/2018/08/Rare-and-inherited-disease-eligibility-criteria-version-5.2.pdf>

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|---|----|
| R137 Congenital heart disease - microarray | 27 |
| R125 Thoracic aortic aneurysm or dissection | 28 |
| R127 Long QT syndrome | 29 |
| R128 Brugada syndrome and cardiac sodium channel disease | 30 |
| R129 Catecholaminergic polymorphic VT | 31 |
| R130 Short QT syndrome | 32 |
| R131 Hypertrophic cardiomyopathy | 33 |
| R132 Dilated and arrhythmogenic cardiomyopathy | 34 |
| R391 Barth syndrome | 36 |
| R133 Arrhythmogenic right ventricular cardiomyopathy | 37 |
| R135 Paediatric or syndromic cardiomyopathy | 38 |
| R136 Primary lymphoedema | 40 |
| R138 Sudden unexplained death or survivors of a cardiac event | 41 |
| R328 Progressive cardiac conduction disease | 41 |
| R384 Generalised arterial calcification in infancy | 43 |
| R140 Elastin-related phenotypes | 44 |
| R441 Unexplained death in infancy and sudden unexplained death in childhood | 44 |

2.1 Pyruvate

(Advice from Dr Lorna Maddocks - Clinical Bio-Chemist, Mon-Wed 01162586553)

- The laboratory provides a pre-weighed sample bottle containing an acid precipitant, which is needed to be in the laboratory **at LRI within 30 minutes on ice** in order to reweigh on a sensitive a balance which is located at LRI and also centrifuge a.s.a.p.
- **A precise measurement of 1 ml of blood** to be added immediately into the sample bottle (if less than 1 ml – insufficient sample for analysis, if significantly greater than 1 ml – insufficient precipitant present to precipitate all the proteins and the assay result will be unreliable)
- The pyruvate result if only of clinical value when paired with simultaneously lactate measurement and **when the lactate level is persistently raised**.
- The blood lactate to pyruvate (L:P) ratio can be used to distinguish between pyruvate dehydrogenase deficiency and other causes of congenital lactic acidosis. In conjunction with an elevated lactate, an L:P ratio greater than 30 suggests inherited disorders of the respiratory chain complex or tricarboxylic acid cycle disorders. In conjunction with an elevated lactate, an L:P ratio less than 25 suggests a defect in pyruvate metabolism.
- An artifactually high L:P ratio can be observed in acutely ill individuals.

Abnormal concentrations of lactate, pyruvate, and the L:P ratio are not diagnostic for any single disorder and must be interpreted in the context of the individual's clinical presentation and other laboratory studies.

2.2 Muscle biopsy for histochemistry, immunochemistry and electron microscopy and for the investigation of mitochondrial disorders.

See Muscle and Skin Biopsy UHL Childrens Medical Guideline C39/2016

Contacts:

1. Chemical Pathology, Dr Elaine Maddocks; Dr Virginia Lee ext 16553.
2. Histopathology – an advice on additional sample requirements before any muscle biopsy sample is collected. Peter Wells-Jordan, ext 16590.
3. In cases of possible mitochondrial cardiomyopathy, Dr Robert McFarland, Consultant Paediatric Neurologist (robert.mcfarland@ncl.ac.uk), can be contacted via:

Newcastle NCG Rare Mitochondrial Disease Service

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Clinical advice: Dr Robert McFarland: via_bernadette.caygill@nuth.nhs.uk
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Diagnostic Laboratory: Prof Robert Taylor: robert.taylor5@nuth.nhs.uk

2.3 Barth Syndrome Testing:

Routine blood testing: Cardiolipin Analysis

Cardiolipin analysis is temporarily unavailable in the UK. The laboratory at Bristol Royal Infirmary continues however to facilitate testing for Barth syndrome by forwarding samples to the Academic Medical Centre (AMC) in Amsterdam for analysis. Analysis of samples sent via the Bristol laboratory to the AMC are free of charge.

Please take 1-3 ml of whole blood collected into a K-EDTA (full blood count) tube, Monday to Thursday only and send within 24 hours of collection by first class post to the:

Metabolic, Neuroendocrine and Nutrition Laboratory
Department of Clinical Biochemistry
Bristol Royal Infirmary
Bristol
BS2 8HW

Please label the box 'Urgent clinical sample - for immediate delivery to the Metabolic, Neuroendocrine and Nutrition Laboratory'.

Alternatively a blood spot can be sent directly to The Academic Medical Centre (AMC), Amsterdam (see <https://www.amc.nl>).

Please note that samples sent directly to the AMC will incur a charge.

Post mortem testing

If a patient is suspected to have died as a result of Barth Syndrome, cardiolipin analysis can be performed in cultured fibroblasts (skin cells). If fibroblasts are unavailable, please contact Vicki Powers to discuss details on 0117 342 2590 or by email at: victoria.powers@UHBristol.nhs.uk

Genetic testing by TAZ Gene Sequencing

Further information on genetic testing in Bristol, and a proforma for referral of samples are given here.

The information on this page is taken from the NHS Barth Syndrome Service site – for further details please visit [Barth Syndrome Service - Testing](#)

Test for it in the following situations:

- **All boys with DCM, especially neonates/infants with CM**
(several large studies suggest that 3-7% of this cohort will have Barth Syndrome; 70% develop CM during their first year)
- **Any boy with unexplained LVNC or HCM with other features suggestive of Barth Syndrome** (e.g. neutropenia, growth failure, feeding problems, myopathy, lactic acidosis, hypoglycaemia) **or X-linked family history**
- **Fetal tissue (fetal fibroblasts or DNA) from recurrent male stillbirth or single stillbirth with suspicion of cardiomyopathy**

3. Interpretation of Investigations

Interpretation of the results of the tests outlined above relies on an understanding of the causes of cardiomyopathies.

Causes of Hypertrophic Cardiomyopathy

Genetic

Gene mutation not always apparent, but definite heritability

- Sarcomeric/other cardiac genetic abnormalities
 - Sarcomeric protein disease
 - Commonest causes of adult HCM
 - 50-60% have mutations in 1 of 11 sarcomere protein genes
 - Also account for over 50% of idiopathic HCM cases in children
 - Z-disk protein disease
 - Small number of patients
 - Calcium-handling disease
 - Small number of patients

Inborn Errors of Metabolism

- <10% of paediatric HCM
- Most cases of HCM due to IEM are due to Glycogen Storage Disorders

- Can be classified as:
 - Accumulation of Toxins
 - Protein metabolism
 - Amino acidopathies
 - Organic acidopathies
 - Urea Cycle Defects
 - Carbohydrate intolerance
 - Lysosomal storage disorders
 - Mucopolysaccharidoses
 - I, II, III, IV, VII
 - Gangliosidoses
 - GM1 and 2
 - Danon Disease (LAMP-2)
 - Fabry Disease
 - Disorders of energy utilization
 - Fatty Acid Metabolism Defects
 - Carnitine transport defects
 - Systemic
 - Muscle
 - CPT II
 - CAT
 - Fatty Acid Oxidation Defects
 - VLCADD
 - LCHADD
 - Glutaric Acidaemia Type II
 - Carbohydrate utilization defects
 - Glycogen storage disorders
 - GSD II (Actually a lysosomal storage disorder, Pompe)
 - GSD III
 - GSD IX
 - AMP Kinase Disease (PRKAG2)
 - Gluconeogenic disorders
 - Glycogenolytic disorders
 - Mitochondrial disorders
 - Respiratory Chain Enzyme Deficiencies
 - Complexes I - V
 - MELAS
 - MERRF
 - Kearns-Sayre
 - Pyruvate dehydrogenase deficiency (Leigh Disease)
 - Barth Syndrome
 - Senger Syndrome
 - Mitochondrial DNA depletion syndrome
 - Peroxisomal disorders

Neuromuscular conditions

- Friedrich's Ataxia
- Myotonic dystrophy

Malformation Syndromes

- RAS/MAPK Diseases
 - Noonan Syndrome
 - Noonan Syndrome with Multiple Lentiginos (Previously LEOPARD syndrome)
 - Costello Syndrome
 - Beckwith-Wiedemann Syndrome
 - Swyer Syndrome (Pure gonadal dysgenesis)

Other causes

- Obesity
- Infant of Diabetic Mother
- Amyloidosis
- Conditions that mimic HCM
 - Athletic training
 - Steroid-induced LVH

Causes of Dilated Cardiomyopathy

- Idiopathic (66%)
- Known causes (34%)
 - Myocarditis
 - Infectious agent
 - Enterovirus, echovirus, coxsackie, HIV
 - Diphtheria, Rheumatic fever, Chagas Disease
 - Neuromuscular
 - Duchenne Muscular Dystrophy
 - Becker Muscular Dystrophy
 - Emery-Dreifuss Muscular Dystrophy
 - Familial
 - AD
 - AR
 - X-linked
 - IEM
 - Mitochondrial
 - Barth Syndrome
 - Primary carnitine deficiency

Causes of Restrictive Cardiomyopathy

- **Idiopathic 90% of childhood causes**
- Fabry's disease – Decreased lysosomal α -galactosidase A in WBC. Pain hands/feet, papules on waist/buttocks
- Gaucher's disease – Hepatosplenomegaly/ Hypersplenism/ Bone marrow infiltrate, flask deformity of distal femur, decreased beta glucosidase in WBC/Skin fibroblasts
- Haemochromatosis- Diabetes/ Bronze skin/ Cirrhosis/ Abdominal pain/ hypogonadism/ : Raised Iron, Ferritin, Increased transferrin saturations > 62%. Do liver biopsy
- Glycogen storage disorder : Lactate/Glucose/ raised TGL/ Raised cholesterol
- Hypereosinophilic (Loeffler) Syndrome : Blood film , raised eosinophil count

- Carcinoid : In appendix. Do 5 HT
- Metastatic malignancy
- Pseudoxanthoma elasticum : Yellow papules and plaques / Eyes- angioid streaks
- Scleroderma- ANA (Anti Scl 70 specificity), Skin signs
- Amyloidosis- in childhood sec to other inflammatory process, Low voltage QRS, Thickened myocardium (sparkling echogenicity)
- Sarcoidosis – Eyes/ Hepatosplenomegaly / erthema nodosum/ polyarthritis/ lungs/ thoracic L.N – Serum ACE level, T cells in BAL

Table 2 Interpretation of Investigations

| 1 st line screening tests | Interpretation |
|---|--|
| Blood | |
| Full blood count | Neutrophils low in Barth Syndrome |
| Urea, creatinine, electrolytes | Allows calculation of anion gap |
| Liver function tests, including bilirubin | Elevated transaminases in fatty acid disorders, GSD II, III May be raised or normal in virtually all other disorders |
| Coagulation screen | Abnormal in hepatic dysfunction associated with e.g. Lysosomal storage disorders |
| Blood gas <i>To allow calculation of anion gap</i> | pH, glucose, lactate Allows assessment of metabolic acidosis <i>Primary metabolic acidosis always present in organic acidaemia, usually present in mitochondrial disorder and fatty acid disorder</i> Allows calculation of anion gap |
| Blood glucose | Hypoglycaemia in <i>Some mitochondrial disorders associated with diabetes</i> <i>Fatty acid metabolism defects</i> <i>GSD III and IX (phosphorylase kinase defect)</i> |
| Ketones | Raised in GSD III, can be raised in organic acidaemias, low or absent in fatty acid disorders Can be raised in unwell child – <i>non-diagnostic pattern on urinary organic acid analysis</i> |
| Bicarbonate | Assessment of metabolic acidosis |
| Ammonia | Raised in Urea cycle defects, organic acidaemias, fatty acid defects, May be raised in aminoacidopathies, mitochondrial disorders |
| Lactate | Sometimes elevated in mitochondrial disorders With abnormal organic acids, suggests fatty acid disorder Raised in Barth syndrome, Senger's Syndrome Allows calculation of lactate:pyruvate ratio – <i>elevated in respiratory chain defects, normal in e.g. pyruvate dehydrogenase deficiency, Normal is <15:1, high is >25:1</i> |
| Pyruvate | Normal or low in respiratory chain defects Elevated in pyruvate metabolism disorders |
| Creatine kinase | Raised in GSD II |
| Free fatty acids | Fatty acid metabolism defects Plasma FFA:beta hydroxybutyrate (intermediate metabolites) |
| Cholesterol | Smith-Lemli-Opitz |
| Amino Acids | Amino acidaemias |
| Acylcarnitine profile | Carnitine transport defects Fatty acid metabolism |
| Thyroid function | Exclude hypothyroidism |
| Blood cultures | |
| Viral PCR <i>Enterovirus, echovirus, coxsackie virus, consider HIV</i> | Myocarditis |
| ASOT <i>Rheumatic fever</i> | Rheumatic fever |
| Vitamin D | Vitamin D deficiency |
| Selenium | Nutritional deficiencies |
| Thiamine (Vit B1) | Nutritional deficiencies |
| | |
| Urine | |
| Amino acids | Elevated in amino acidaemias |
| Organic acids | Abnormal in organic acidaemias, fatty acid oxidation defects, Barth Syndrome |
| Oligosaccharides | Raised in GSD II, other lysosomal disorders |

| | |
|--|--|
| Glycosaminoglycans | Elevated in MPS, lysosomal disorders |
| Ketones | Elevated in organic acidaemias |
| Glucose | Exclude DM, abnormal in GSD's (some) |
| Reducing substances | Galactosaemia |
| 2nd Line Testing | |
| Muscle and Skin Biopsy for mitochondrial disorders | |
| Histology | |
| Electron microscopy | |
| Enzyme assays | For respiratory chain enzymes, Complexes I to V |
| Mitochondrial DNA analysis | For specific mitochondrial DNA mutations, may also look at nuclear DNA |
| Brain MRI | |
| Suspected mitochondrial disorders | May see basal ganglia changes, general cortical changes |
| Blood | |
| Cardiolipin Levels (Barth Syndrome) | |
| White cell enzymes <i>For lysosomal storage disorders (MOS II & III), if abnormal urinary oligosaccharides, GAG's, elevated CK or ALT/AST</i> | Specific enzyme assays |
| Transferrin and Apolipoprotein isoforms <i>if Congenital Disorder of Glycosylation suspected</i> | Abnormal in CDG Type I or II respectively |
| Phytanic Acid <i>if Refsum Disease suspected</i> | |

Anion gap raised in

- organic acidaemias,
- fatty acid oxidation defects,
- respiratory chain disorders,
- pyruvate disorders

Differentiating requires assessment of lactate and pyruvate levels

4. Education and Training

No training required for this guideline

5. Monitoring Compliance

None currently identified

6. Supporting References

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4. Moak, J. P. and Kaski, J. P. Hypertrophic cardiomyopathy in children. *Heart*. 2012; 98:1044-1054
5. Sutton, V. R. Inborn errors of metabolism. *UpToDate*. 2012. Accessed April 2013
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7. Key Words

Barth syndrome, Biopsy, Cardiomyopathy, Dilated cardiomyopathy, Hypertrophic cardiomyopathy, Inborn errors of metabolism, Mitochondrial disorders, Muscle biopsy Myopathy, Restrictive cardiomyopathy, Skin Biopsy

The Trust recognises the diversity of the local community it serves. Our aim therefore is to provide a safe environment free from discrimination and treat all individuals fairly with dignity and appropriately according to their needs. As part of its development, this policy and its impact on equality have been reviewed and no detriment was identified.

| CONTACT AND REVIEW DETAILS | |
|--|--------------------------------------|
| Guideline Lead (Name and Title) Simon Chiles – Advanced Nurse Practitioner Suhair Shebani - Consultant | Executive Lead Chief Nurse |
| Details of Changes made during review: August 2023: updated muscle biopsy and Barth Syndrome and shortened the document | |